

Effects of Root Canal Irrigants and Medicaments on Dentin and Vice Versa: A Review of Literature

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Abstract

Microorganisms are very important in the initiation of pulpo-periapical pathosis. Due to the complexity of the root canal system, the mechanical instrumentation of the root canal system should be supplemented with proper canal irrigants and medicaments. It has been revealed that due to the interactions of canal irrigants and medicaments with dentin and contents of the canal system, the antimicrobial effect of canal irrigants and medicaments in canal (*ex vivo*) are different from that *in vitro*. Furthermore, root canal irrigants may decrease the fracture resistance of dentin. The purpose of this paper was to review the data on the interactions between root canal irrigants/medicaments and dentin/root canal contents.

Keywords: Calcium hydroxide, Chlorhexidine, Dentin, EDTA, Iodine compounds, MTA, MTAD, NaOCl

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Introduction

Microorganisms play an important role in initiation of pulpo-periapical pathosis (1-3). Elimination of microorganisms from an infected root canal system (RCS) is a complicated stage of endodontic treatment. Numerous measures such as usage of different mechanical instrumentation techniques, irrigation solutions, and intra-canal medicaments have been proposed to decrease the number of microorganisms from the RCS. Mechanical preparation alone cannot predictably result in bacteria-free RCS, which is not surprising given the complexity of the RCS. On the contrary, there is both clinical and *in vitro* documents indicating that mechanical preparation may leave significant areas of the RCS untouched (4-6). Any pulp tissue left in the RCS may serve as a nutrient source for microorganisms. This tissue may be digested by the bacteria up to two months, depending on whether the RCS is open to the oral environment or not (7). Furthermore, tissue remnants may impede the antibacterial effects of irrigants/medicaments. Therefore, some forms of medicaments and irrigants may be necessary to remove debris and tissue from the RCS and kill the remaining microbes. The aim of the present review was to collect data on the effects of root canal irrigation solutions and intra-canal medicaments on dentin by reviewing the related articles.

Dentin Chemistry

As the main hard tissue of the tooth structure, dentin consists of dentinal tubules surrounded by highly mineralized peri-tubular and inter-tubular dentin. Depending on the tooth location and age, its composition may show some variations. Furthermore, external irritation, such as caries, may affect dentin composition (8). Amongst the hard tissues of the tooth, dentin is chemically the closest tissue to the bone (9). Its inorganic components are composed of calcium phosphate compounds, mostly apatite. In addition, small amounts of potassium, sodium, magnesium, carbonate, chloride, and fluoride may be found in dentin (10). Type I collagen is its major organic component, although small amounts of type III and V have been found, too (9, 11). Type I collagen has higher proportions of hydroxylysine in dentin (11). As a result, it has more intra-molecular/intermolecular cross-links than type I collagen in bone. Cross-links increase the structural stability and strength of dentin collagen. They also contribute to making dentin collagen relatively insoluble in acid as compared with collagen from other sources. Consequently, acid etch of dentin removes the great part of mineral phase, whereas the collagen may remain almost intact (9, 11).

Inorganic component of dentin (Apatite)

Non-homogeneous apatite form most of the mineral content of dentin. Based on spectroscopy analysis, LeGeros (12) showed that the calcium content of dentin apatite is less than that of enamel, whereas the amount of carbonated apatite was slightly more in dentin than in enamel (13). Furthermore, according to Tsuda et al. (14), the concentration of HPO_4^{2-} is higher and the concentration of OH^- is lower in dentin, comparing the enamel. Hydroxyapatite is surrounded by a layer of adsorbed ions and water. The hydration layer and exchange/adsorption of ions allow changes in the chemical microenvironment, reflecting pH and interaction with chemical compounds (10).

Buffering effect of dentin

Bone apatite is a major carbonate reservoir, and so providing buffering for all acid-base disturbances and maintaining its balance in the body (15-17). Considering a quite similar chemical composition between the bone and dentin, it can be expected that dentin can possess a corresponding buffering effect on bases and acids. According to Wang and Hume (18), the buffering efficacy of dentin against alkali is weaker but considerable. Dentin chips are able to keep the pH unchanged after the addition of NaOH or HCl. Inorganic apatite is the main responsible for the buffering effect of dentin. However, other inorganic and even organic components may also contribute to the buffering effect. (15, 17, 18).

Sodium hypochlorite (NaOCl)

NaOCl acts as a solvent for organic and fatty acids, by transforming them into fatty acid salts and glycerol reducing the surface tension of the solution (16). It neutralizes amino-acids forming water and salt. With the exit of hydroxyl ions, there is a reduction in pH. In contact with organic tissues, hypochlorous acid which is present in NaOCl solution, acts as a solvent and releases chlorine, which combined with the protein amino group, forms chloramines that interfere in cell metabolism. Hypochlorous acid and hypochlorite ions lead to amino-acid hydrolysis and degradation (19). As a strong oxidant, chlorine presents antibacterial actions inhibiting bacterial enzymes, thereby leading to the irreversible oxidation of sulphhydryl group of bacterial enzymes (19, 20). Considering the physic-chemical properties of NaOCl when in contact with tissues, these reactions may be verified. Its antibacterial mechanism of action can verify its physic-chemical characteristics and reaction with tissues (16).

The antibacterial effectiveness of NaOCl, based on its pH is similar to the mechanism of calcium hydroxide (CH) (20). The high pH of NaOCl interferes in the integrity of cytoplasmic membrane with enzymatic inhibition, and phospholipid degradation. The amino-acid chloramination reaction forming chloramines interferes with cell metabolism. This enzyme inactivation may be observed in the reaction of chlorine with amino groups and oxidation of sulphhydryl groups of bacterial enzymes (19, 20). Therefore, NaOCl presents antibacterial activity with acting on microbial enzymatic sites promoting inactivation originated by chloramination action. Dissolution of organic tissues can be verified in the saponification reaction when NaOCl degrades lipids (20).

Buffering effect of dentin on the antibacterial activity of NaOCl

The organic components of dentine alone accounted for 1.5% of the total buffering capacity (21, 22). The RCS milieu is mixture of a variety of inorganic and organic compounds. Hydroxyapatite which is the principal part of dentine is the major representative of the inorganic base. Difficulty in designing studies rendering reliable data is a great challenge for many years. Ultimately, Haapasalo et al. (23) introduced a new dentine powder for investigating the inhibitory effect of dentine on RCS irrigants and also medicaments. NaOCl which is a non-specific oxidizer, with amino-acids by chloramination and neutralization reactions, leading to amino-acids degradation (19, 20). An immuno-histochemical research showed that type I collagen and glycosaminoglycan lost their immune-reactivity after NaOCl usage when a dematerialized dentine was used (24). However, in intact dentine model, this was minimal, proposing that hydroxyapatite has a protective role by embedding

proteins such as collagen against the oxidation of NaOCl. Haapasalo et al. (23) have showed the inhibitory effect of dentine on the effectiveness of 1% NaOCl against *Enterococcus faecalis*. They also showed that killing all of the microorganisms required 24 hours of incubation with hypochlorite. However, after one hour of incubation, all of the microorganisms were still viable. It seems that dentine decreases or inhibits the antimicrobial effect of NaOCl.

Effect of NaOCl on the composition and structure of dentine

The impact of NaOCl on the dentine matrix is one of its side effects. Dentine is composed of 22% organic component specially type I collagen, which contributes to dentine mechanical properties. It can fragment long peptide chains and chlorinate protein terminal groups (25). Consequently, it affects dentine mechanical properties by the degradation of organic dentine materials (26). It has been shown that concentrated hypochlorite solutions may cause untoward effects on dentine biomechanical properties (27). A 2-hour dentine exposure to $\geq 3\%$ NaOCl can reduce the elastic modulus and flexural strength of dentine (28, 29). Mountouris et al. (30) found that both 5% NaOCl solution and acid-etched coronal dentine surfaces reduced the organic matrix but did not affect carbonates and phosphates. Di Renzo et al. (31) showed that treatment with NaOCl using a photo-acoustic FTIRS technique, induced a slow and heterogeneous removal of its organic phase, leaving calcium hydroxyapatite and carbonate apatite unchanged. Another study showed that 5% NaOCl induced alterations in dentine collagen, whereas hydroxyapatite showed a protective role in stability of organic matrix. Marending et al. (26) found that NaOCl caused a concentration-dependent reduction of flexural strength and elastic modulus in dentine. The carbon and nitrogen content were significantly decreased.

Chlorhexidine (CHX)

Structure and mechanism of action

CHX is a synthetic cationic bis-biguanide. It is a positively charged hydrophobic/lipophilic molecule interacts with phospholipids on the bacterial cell membrane (32, 33). Its efficacy is due to the interaction of the positive charge of the molecule and the negatively charged phosphate groups on bacterial cell walls (34, 35). This enhances the cell wall permeability, which allows the CHX molecule to penetrate into the microorganism. The most common oral form of CHX is CHX gluconate which is water-soluble that dissociates and releases the positively charged CHX component at physiologic pH (32, 33, 36). At low concentration (0.2%), potassium and phosphorous can leak out of the cell (35).

Effect of CHX on dentin

CHX significantly decreases the micro-hardness of root dentin at 500 and 1000 μm from the pulpo-dentinal

junction (37). Also, it has been shown that 2% CHX gel cannot adversely affect dentin micro-hardness when associated with the bleaching agents (38). Ari et al. (39) showed that all widely used irrigation solutions except for 0.2% CHX, significantly reduced the micro-hardness of root dentin. In this study, 3% H_2O_2 and 0.2% CHX gluconate had no effect on root dentin roughness. According to the results of this study, 0.2% CHX gluconate may be an appropriate irrigation solution in endodontic treatment because of its weak effect on the micro-hardness and roughness of dentin. In another study, 0.2% CHX was reported to have no great effect on dentin micro-hardness (40). Aslantas et al. (41) also studied the effects of endodontic irrigants on dentin micro-hardness in presence of surface-modifying agents (17% ethylenediaminetetraacetic acid (EDTA), REDTA, 2% CHX, 2% CHX with surface modifiers (CHX-Plus), 6% NaOCl, or 6% NaOCl with surface modifiers (Chlor-XTRA). They showed that EDTA, REDTA, NaOCl, and Chlor-XTRA significantly decreased dentin micro-hardness. Marcelino et al. (42) concluded that dentin micro-hardness was decreased after exposure to CHX, NaOCl, phosphoric acid, and sodium ascorbate. However, dentin flexural strength was not affected by the chemical agents. Kara Tuncer et al. (43) also showed that maleic acid significantly reduced dentin micro-hardness compared to EDTA+CHX, EDTA+NaOCl, QMix. Das et al. (44) showed that NaOCl+Q Mix were least detrimental to dentin micro-hardness comparing *Morinda citrifolia* juice and conventional irrigation solutions.

Modulating effect of dentine on CHX

Portenier et al. (45) evaluated the inhibition of the antibacterial effect of saturated CH solution, CHX acetate, and IKI by dentine, hydroxyapatite, and bovine serum albumin. They concluded that 0.05% CHX was inhibited by bovine serum albumin and slowed down by dentine. However, hydroxyapatite had little inhibitory effect on CHX. They also showed that inorganic hydroxyapatite had little or no inhibitory effect against CHX comparing dentine, whereas bovine serum albumin was the strongest inhibitor of CHX. Portenier et al. (46) also concluded that dentine pre-treated by citric acid or EDTA showed only slight inhibition dentine matrix, whereas killed microbial cells were the most effective inhibitors of CHX. Another study demonstrated that the presence of dentine or bovine serum albumin caused delay in killing of *Enterococcus faecalis* by CHX (47). The inhibitory effect of bovine serum albumin on antimicrobial effect of CHX was confirmed by Sassone et al. (48). It seems that dentine and its components (hydroxyapatite and collagen), killed microorganisms, and inflammatory exudates in the RCS can reduce or inhibit the antimicrobial activity of irrigants (such as CHX) and medicaments.

MTAD

BioPure (MTAD) (Dentsply, Tulsa Dental, Tulsa, OK, USA), introduced by Torabinejad and Johnson, is a mixture of 3% doxycycline, 4.25% citric acid, and a detergent (Polysorbate 80) (49).

MTAD can remove the smear layer (49) and affect *Enterococcus faecalis* (50-52). Shabahang et al. (51) compared the antimicrobial effect of a combination of 1.3% NaOCl as an irrigant and MTAD as a final rinse with that of 5.25% NaOCl. They concluded that the first combination is more effective in RCS disinfection than using 5.25% NaOCl alone. However, Tay et al. (53) found that when MTAD was applied to 1.3% NaOCl-irrigated dentine, its antibacterial substantivity was decreased.

Effect of MTAD on dentin

Machnick et al. (54) showed no significant difference in modulus of elasticity and flexural strength between dentin exposed to saline and MTAD. Dineshkumar et al. (55) evaluated the effect of 17% EDTA, MTAD, and 18% HEBP on dentin micro-hardness. MTAD showed the highest effect on dentin micro-hardness. On the other hand, Ulusoy and Gurgol (56) demonstrated that MTAD had the least effect on dentin micro-hardness. In another study, MTAD significantly reduced dentin micro-hardness (57).

Effect of dentin on MTAD

A recent study confirmed earlier findings about the inhibition of CHX activity by dentin. They also showed a corresponding inhibition of MTAD by dentin (58). Although MTAD killed *Enterococcus faecalis* within 5 minutes, the addition of dentin slowed down the killing process (58).

Iodine compounds

Iodine is bactericidal, virucidal, fungicidal, tuberculocidal, and sporicidal by attack the proteins. Aqueous iodine solutions are unstable, with molecular iodine being responsible for the antimicrobial activity. Iodophors are complexes of iodine and a solubilizing agent or carrier (59).

Buffering effect of dentin

The presence of dentin is responsible for inhibitory patterns on the activity of iodine solutions. Haapasalo et al. (34) demonstrated that dentin powder effectively abolished the effect of 0.2% IKI 0.4%, it took only 5 minutes to kill *Enterococcus faecalis*. Portenier et al. (45) showed that hydroxyapatite caused little or no inhibition, whilst the dentinal collagen matrix effectively inhibited 0.1% IKI 0.2%.

EDTA

EDTA which is a colorless, water-soluble solid is widely used as a chelating agent. It binds to metals through 4 carboxylate groups and 2 amine groups. It forms especially strong complexes with Mn, Cu, Fe, and Co. It is mostly synthesized from ethylenediamine,

formaldehyde, sodium cyanide, and water. After being bound by EDTA, metal ions remain in solution but exhibit diminished reactivity (60-62).

Effects on dentine micro-hardness

Chelators can decrease dentine micro-hardness, whereby the greatest differences are to be found in dentine immediately adjacent to the canal (63). The effect of the chelator is already apparent after 5 minutes. Cruz-Filho et al. (64) showed that EDTA and citric acid had the greatest effect on dentin micro-hardness. In another study, Ballal et al. (65) found no significant difference between EDTA and maleic acid in reduction of dentine micro-hardness.

Eldeniz et al. (66) assessed the effect of EDTA and citric acid on dentine micro-hardness and roughness and showed a significant difference in micro-hardness among the test groups, citric acid group being the least hard. In addition, Ari et al. (67) and Cruz-Filho et al. (68) confirmed the decrease of dentine micro-hardness after using EDTA. De-Deus et al. (69) assessed the effect of EDTA, EDTAC, and citric acid on dentine micro-hardness and found that micro-hardness decreased with the increasing time of the application of chelators.

CH

CH was originally introduced as a pulp capping agent (70, 71). It has low solubility in water, which decreases by temperature rise (71). The dissociation coefficient of CH permits a slow release of both hydroxyl ions and calcium. The low solubility is a suitable characteristic because a long period is necessary for CH to become soluble in tissue fluids when in contact with vital tissues (70, 72).

Buffering effect of dentine on CH

Haapasalo et al. (23) showed that dentine powder effectively abolished killing of *Enterococcus faecalis* by CH. On the other hand, in absence of dentine, saturated CH killed *Enterococcus faecalis* in a few minutes. Portenier et al. (45) demonstrated that hydroxyapatite had an effect similar to that of dentine on CH. The substantial effect of dentine on the antimicrobial effect of CH may be attributed to the buffering action of dentine against alkali (18). Buffering by dentine may be the main factor behind the decreased antimicrobial effect of CH. It is possible that outside the canal, CH is present at concentrations even below that level (23). Besides dentine, the remnants of necrotic pulp and inflammatory exudative fluid may affect the antimicrobial effect of disinfectants (8).

Effect of CH on dentine

Endodontic treatment of immature teeth with non-vital pulp is a great challenge. Apexification by CH may induce apical healing through induction of an apical barrier whilst at the same time; the high pH provides an antimicrobial activity. The flexural strength of dentine may depend on the link between the hydroxyapatite

crystals and the collagenous network. The organic matrix is composed of acid proteins and proteoglycans (73-75). These substances act as bonding agents between the hydroxyapatite crystals and collagen network (75). Rosenberg et al. (76) showed that the micro-tensile fracture strength of teeth and was decreased by almost 50% following 7-84 days. Another study showed this to be 32% (77). The results of another study indicated that the fracture strength of sheep dentine was decreased by 50% following CH treatment after one year (78). Kawamoto et al. (79) have also concluded that CH paste may significantly increase the elastic modulus of dentine. Grigoratos et al. (80) reported that CH may cause reduction in flexural strength of dentine. Andreasen et al. (75) concluded that the fracture strength of CH-filled immature teeth was halved in 1 year and attributed the frequent reports of the fractures of open apex teeth filled with CH to extended exposure periods. Doyon et al. (81) examined the resistance of dentine to CH and found that the fracture resistance of dentine was reduced after six months.

Mineral trioxide aggregate (MTA)

MTA is a mixture of refined Portland cement, bismuth oxide, and trace amounts of SiO₂, CaO, K₂SO₄, and Na₂SO₄ (82). MTA powder is mixed with supplied sterile water. Upon hydration, MTA materials form a colloidal gel that solidifies to a hard structure in approximately 3-4 hours, with moisture from the surrounding tissues (83). Hydrated MTA has an initial pH of 10.2, which rises to 12.5 about 3 hours after mixing (84). The compressive strength of MTA increases in the presence of moisture for up to 21 days, while MTA micro-hardness and hydration behavior adversely affected with exposure to the pH range of inflammatory condition as compared to physiologic environment (85).

MTA and susceptibility to root fracture

The most promising alternative to long-term CH therapy for induction of apexification is the use of MTA as an apical barrier or apical plug (83). However, the increase of root resistance to fracture remains a challenge (84, 85). Andreasen et al. (86) showed that when CH was kept in the canals of immature teeth for 1 month, followed by MTA filling, there was no significant reduction in the fracture strength within 100 days. Bortoluzzi et al. (87) showed that the combined use of MTA and metallic posts increased the resistance to the fracture. Hatibović-Kofman et al. (88) showed that fracture strength of teeth treated with CH or MTA decreased but not significantly over time. For the MTA-treated teeth, the fracture strengths were not significantly different between the untreated and CH-treated teeth after 15 days or 60 days. Tuna et al. (89) assessed the fracture resistance of immature teeth filled with BioAggregate (BA), MTA, and CH *in vitro* and showed that BA group exhibited the highest fracture resistance,

while CH group showed the lowest amount of resistance. Aksel et al. (90) also showed that MTA increased the resistance of immature teeth to vertical fracture. Guven et al. (91) assessed fracture resistance of immature teeth filled with BA, MTA, and EndoSequence Root Repair Material. The results revealed that BA-filled teeth had higher fracture resistance than other groups after 24 months. Schmoldt et al. (92) showed that gutta-percha or MTA cannot increase the fracture resistance. Likewise, Sawyer et al. (93) showed that the flexural strength of dentin exposed to MTA Plus decreased after 3 months. Elnaghy and Elsaka (94) assessed the fracture resistance of immature teeth filled with BD and white MTA. They observed that after 1 year, there was no difference between these materials. The results of another study showed that one year after exposure to CH and MTA, the flexural strength of dentin reduced to 72% and 39%, respectively (95). Forghani et al. (96) showed that MTA increased the fracture resistance of dentin, while Portland cement had no effect on dentin fracture resistance.

References

1. Kakehashi S, Stanley HR, Fitzgerald RJ. The effects of surgical exposure of dental pulps in germ-free and conventional laboratory rats. *Oral Surg Oral Med Oral Pathol.* 1965;18:340-348.
2. Möller AJ, Fabricius L, Dahlen G, et al. Influence on periapical tissues of indigenous oral bacteria and necrotic pulp tissue in monkeys. *Scand J Dent Res.* 1981;89(6):475-484.
3. Sundqvist G. Ecology of the root canal flora. *J Endod.* 1992;18(9):427-430.
4. Bystrom A, Sundqvist G. Bacteriologic evaluation of the efficacy of mechanical root canal instrumentation in endodontic therapy. *Scand J Dent Res.* 1981;89(4):321-328.
5. Hess, W. Anatomy of root canals of the teeth of the permanent dentition. London: Sons and Danielson Ltd, 1925.
6. Peters OA, Laib A, Gohring TN, et al. Changes in root canal geometry after preparation assessed by high resolution computed tomography. *J Endod.* 2001;27(1):1-6.
7. Zehnder M. Root canal irrigants. *J Endod.* 2006;32(5):389-398.
8. Gabor C, Tam E, Shen Y, Haapasalo M. Prevalence of internal inflammatory root resorption. *J Endod* 2012;38(1):24-27.
9. Nanci A. Ten Cate's oral histology. St Louis: Mosby, 2003.
10. Jenkins G. The physiology and biochemistry of the mouth. Oxford: Blackwell Scientific Publications, 1978.
11. Gage J, Francis M, Triffitt J. Collagen and dental matrices. Boston: Butterworth & Co. Ltd, 1989.

12. LeGeros RZ. Calcium phosphates in oral biology and medicine. *Monogr Oral Sci.* 1991;15(1):1–201.
13. LeGeros RZ, Kijkowska R, Bautista C, LeGeros JP. Synergistic effects of magnesium and carbonate on properties of biological and synthetic apatites. *Connect Tissue Res.* 1995;33(1-3):203–209.
14. Tsuda H, Ruben J, Arends J. Raman spectra of human dentin mineral. *Eur J Oral Sci.* 1996;104(2):123–131.
15. Poyart CF, Bursaux E, Freminet A. The bone CO₂ compartment: evidence for a bicarbonate pool. *Respir Physiol.* 1975;25(1):89–99.
16. Bettice JA, Gamble JL Jr. Skeletal buffering of acute metabolic acidosis. *Am J Physiol.* 1975;229(6):1618–1624.
17. Lemann J Jr, Litzow JR, Lennon EJ. The effects of chronic acid loads in normal man: further evidence for the participation of bone mineral in the defense against chronic metabolic acidosis. *J Clin Invest.* 1966;45(10):1608–1614.
18. Wang JD, Hume WR. Diffusion of hydrogen ion and hydroxyl ion from various sources through dentine. *Int Endod J.* 1988;21(1):17–26.
19. Mohammadi Z. NaOCl in endodontics: an update review. *Int Dent J.* 2008;58(6):329–341.
20. Mohammadi Z, Shalavi S. Antimicrobial activity of NaOCl in endodontics. *J Mass Dent Soc.* 2013;62(1):28–31.
21. Haapasalo M, Qian W, Portenier, I et al. Effects of dentin on the antimicrobial properties of endodontic medicaments. *J Endod.* 2007;33(8):917–925.
22. Camps J, Pashley DH. Buffering action of human dentine *in vitro*. *J Adhes Dent.* 2000;2(1):39–50.
23. Haapasalo HK, Siren EK, Waltimo TM. Inactivation of local root canal medicaments by dentine: an *in vitro* study. *Int Endod J.* 2000;33(2):126–131.
24. Oyarzun A, Cordero AM, Whittle M. Immunohistochemical evaluation of the effects of NaOCl on dentine collagen and glycosaminoglycans. *J Endod.* 2002;28(3):152–156.
25. Stoward PJ. A histochemical study of the apparent deamination of proteins by NaOCl. *Histochem.* 1975;45(3):213–226.
26. Marending M, Luder UH, Brunner TJ, et al. Effect of NaOCl on human root dentine – mechanical, chemical and structural evaluation. *Int Endod J.* 2007;40(10):786–793.
27. Slutzky-Goldberg I, Maree M, Liberman R, et al. Effect of NaOCl on dentin microhardness. *J Endod.* 2004;30(12):880–882.
28. Grigoratos D, Knowles J, Ng YL, et al. Effect of exposing dentine to NaOCl and calcium hydroxide on its flexural strength and elastic modulus. *Int Endod J.* 2001;34(2):113–119.
29. Sim TP, Knowles JC, Ng YL, et al. Effect of NaOCl on mechanical properties of dentine and tooth surface strain. *Int Endod J.* 2001;34(2):120–132.
30. Mountouris G, Silikas N, Eliades G. Effect of NaOCl treatment on the molecular composition and morphology of human coronal dentin. *J Adhes Dent.* 2004;6(3):175–182.
31. Di Renzo M, Ellis TH, Sacher E, et al. A photoacoustic FTIRS study of the chemical modifications of human dentin surfaces: II. Deproteinization. *Biomaterials.* 2001; 22(8):793–797.
32. Mohammadi Z, Abbott PV. The properties and applications of chlorhexidine in endodontics. *Int Endod J.* 2009;42(4):288–302.
33. Mohammadi Z, Jafarzadeh H, Shalavi S. Antimicrobial efficacy of chlorhexidine as a root canal irrigant: a literature review. *J Oral Sci.* 2014;56(2):99–103.
34. Gomes BPFA, Souza SFC, Ferraz CCR et al. Effectiveness of 2% chlorhexidine gel and calcium hydroxide against *Enterococcus faecalis* in bovine root dentine *in vitro*. *Int Endod J.* 2003;36(4):267–275.
35. Gomes BP, Sato E, Ferraz CC, Teixeira FB, Zaia AA, Souza-Filho FJ. Evaluation of time required for recontamination of coronally sealed canals medicated with calcium hydroxide and chlorhexidine. *Int Endod J.* 2003;36(9):604–609.
36. Greenstein G, Berman C, Jaffin R. Chlorhexidine: an adjunct to periodontal therapy. *J Periodontol.* 1986;57(6):370–376.
37. Oliveira LD, Carvalho CA, Nunes W, Valera MC, Camargo CH, Jorge AO. Effects of chlorhexidine and NaOCl on the microhardness of root canal dentin. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2007;104(4):e125–e128.
38. de Oliveira DP, Teixeira EC, Ferraz CC, Teixeira FB. Effect of intracoronal bleaching agents on dentin microhardness. *J Endod.* 2007;33(4):460–462.
39. Ari H, Erdemir A, Belli S. Evaluation of the effect of endodontic irrigation solutions on the microhardness and the roughness of root canal dentin. *J Endod.* 2004;30(11):792–795.
40. Patil CR, Uppin V. Effect of endodontic irrigating solutions on the microhardness and roughness of root canal dentin: an *in vitro* study. *Indian J Dent Res.* 2011;22(1):22–27.
41. Aslantas EE, Buzoglu HD, Altundasar E, Serper A. Effect of EDTA, NaOCl, and chlorhexidine gluconate with or without surface modifiers on dentin microhardness. *J Endod.* 2014;40(6):876–879.
42. Marcelino AP, Bruniera JF, Rached-Junior FA, Silva SR, Messias DC. Impact of chemical agents for surface treatments on microhardness and flexural strength of root dentin. *Braz Oral Res.* 2014;28.

43. Kara Tuncer A, Tuncer S, Siso SH. Effect of QMix irrigant on the microhardness of root canal dentin. *Aust Dent J.* 2015;60(2):163-168.
44. Das A, Kottoor J, Mathew J, Kumar S, George S. Dentin microhardness changes following conventional and alternate irrigation regimens: an *in vitro* study. *J Conserv Dent.* 2014;17(6):546-549.
45. Portenier I, Haapasalo H, Rye A, Waltimo T, Orstavik D, Haapasalo M. Inactivation of root canal medicaments by dentine, hydroxylapatite and bovine serum albumin. *Int Endod J.* 2001;34(3):184-188.
46. Portenier I, Haapasalo H, Ørstavik D, Yamauchi M, Haapasalo M. Inactivation of the antibacterial activity of iodine potassium iodide and chlorhexidine digluconate against *Enterococcus faecalis* by dentin, dentin matrix, type-I collagen, and heatkilled microbial whole cells. *J Endod.* 2002;28(9):634-637.
47. Portenier I, Waltimo T, Ørstavik D, Haapasalo M. Killing of *Enterococcus faecalis* by MTAD and chlorhexidine digluconate with or without cetrimide in the presence or absence of dentine powder or BSA. *J Endod.* 2006;32(2):138-141.
48. Sassone LM, Fidel R, Fidel S, Vieira M, Hirata R Jr. The influence of organic load on the antimicrobial activity of different concentrations of NaOCl and chlorhexidine *in vitro*. *Int Endod J.* 2003;36(12):848 – 852.
49. Torabinejad M, Johnson WB. Irrigation solution and methods of use. Philadelphia, PA, USA: US Patent & Trade Mark Office. United States Patent Application 20030235804; December 25.
50. Shabahang S, Torabinejad M. Effect of MTAD on *Enterococcus faecalis*-contaminated root canals of extracted human teeth. *J Endod.* 2003;29(9):576-579.
51. Shabahang S, Pouresmail M, Torabinejad M. *In vitro* antibacterial efficacy of MTAD and NaOCl. *J Endod.* 2003;29(7):450-452.
52. Torabinejad M, Shabahang S, Apécio R, Kettering JD. The antimicrobial effect of MTAD: an *in vitro* investigation. *J Endod.* 2003;29(6):400-413.
53. Tay FR, Hiraishi N, Schuster GS, et al. Reduction in antimicrobial substantivity of MTAD after initial NaOCl irrigation. *J Endod.* 2006;32(10):970-975.
54. Machnick TK, Torabinejad M, Munoz CA, Shabahang S, Macknick T. Effect of MTAD on flexural strength and modulus of elasticity of dentin. *J Endod.* 2003;29(11):747-450.
55. Dineshkumar MK, Vinothkumar TS, Arathi G, Shanthisree P, Kandaswamy D. Effect of ethylene diamine tetra-acetic acid, MTAD™, and HEBP as a final rinse on the microhardness of root dentin. *J Conserv Dent.* 2012;15(2):170-173.
56. Ulusoy Öİ, Görgül G. Effects of different irrigation solutions on root dentine microhardness, smear layer removal and erosion. *Aust Endod J.* 2013;39(2):66-72.
57. Aranda-Garcia AJ, Kuga MC, Chavez-Andrade GM, et al. Effect of final irrigation protocols on microhardness and erosion of root canal dentin. *Micros Res Tech.* 2013;76(10):1079-1083.
58. Portenier I, Waltimo T, Ørstavik D, Haapasalo M. Killing of *Enterococcus faecalis* by MTAD and chlorhexidine digluconate with or without cetrimide in the presence or absence of dentine powder or BSA. *J Endod.* 2006;32(2):138-141.
59. Mohammadi Z. Iodine compounds in endodontics: an update review. *Dent Today.* 2009;28(6):58, 60-63.
60. Mohammadi Z, Shalavi S, Jafarzadeh H. Ethylenediamine tetraacetic acid in endodontics. *Eur J Dent.* 2013;7:S135-142.
61. Holleman AF, Wiberg E. *Inorganic Chemistry.* San Diego: Academic Press, 2001.
62. Harris DC. *Quantitative Chemical Analysis.* New York: W.H. Freeman Company; 2007.
63. Pawlicka H. The use of chelating agents for widening of the root canals. Determination of microhardness. *Stomatol DDR.* 1982;32(5):355-361.
64. Cruz-Filho AM, Sousa-Neto MD, Savioli RN, Silva RG, Vansan LP, Pécora JD. Effect of chelating solutions on the microhardness of root canal lumen dentin. *J Endod.* 2011; 37(3):358-362.
65. Ballal NV, Mala K, Bhat KS. Evaluation of the effect of maleic acid and ethylenediaminetetraacetic acid on the microhardness and surface roughness of human root canal dentin. *J Endod.* 2010;36(8):1385-1388.
66. Eldeniz AU, Erdemir A, Belli S. Effect of EDTA and citric acid solutions on the microhardness and the roughness of human root canal dentin. *J Endod.* 2005;31(2):107-110.
67. Ari H, Erdemir A, Belli S. Evaluation of the effect of endodontic irrigation solutions on the microhardness and the roughness of root canal dentin. *J Endod.* 2004; 30(11):792-795.
68. Cruz-Filho AM, Sousa-Neto MD, Saquy PC, Pécora JD. Evaluation of the effect of EDTAC, CDTA, and EGTA on radicular dentin microhardness. *J Endod.* 2001;27(3):183-184.
69. De-Deus G, Paciornik S, Mauricio MH. Evaluation of the effect of EDTA, EDTAC and citric acid on the microhardness of root dentine. *Int Endod J.* 2006;39(5):401-407.
70. Mohammadi Z, Dummer P. Properties and applications of calcium hydroxide in endodontics and dental traumatology. *Int Endod J.* 2011;44(8):697-730.
71. Mohammadi Z, Shalavi S, Yazdizadeh M. Antibacterial activity of calcium hydroxide in endodontics: a review. *Chonnam Med J.* 2012; 48(7-8):133-140.

72. Farhad A, Mohammadi Z. Calcium hydroxide: a review. *Int Dent J*. 2005;55(5):293-301.
73. Granath L. Hard tissue barrier formation in pulp-tomized monkey teeth capped with cyanoacrylate or calcium hydroxide for 10 and 60 minutes. *J Dent Res*. 1987;66(6):1166-1174.
74. Frank AL. Therapy for the divergent pulpless tooth by continued apical formation. *J Am Dent Assoc*. 1966; 72(1):87-93.
75. Andreasen JO, Farik B, Munksgaard EC. Long-term calcium hydroxide as a root canal dressing may increase risk of root fracture. *Dent Traumatol*. 2002;18(3):134-137.
76. Rosenberg B, Murray PE, Namerow K. The effect of calcium hydroxide root filling on dentin fracture strength. *Dent Traumatol*. 2007;23(1):26-29.
77. White JD, Lacefield WR, Chavers LS, Eleazer PD. The effect of three commonly used endodontic materials on the strength and hardness of root dentine. *J Endod*. 2002;28(12):828-830.
78. Andreasen FM, Andreasen JO, Bayer T. Prognosis of root-fractured permanent incisor-prediction of healing modalities. *Endod Dent Traumatol*. 1989;5(1):11-22.
79. Kawamoto R, Kurokawa H, Takubo C, Shimamura Y, Yoshida T, Miyazaki M. Change in elastic modulus of bovine dentine with exposure to a calcium hydroxide paste. *J Dent*. 2008;36(11):959-964.
80. Grigoratos D, Knowles J, Ng YL, Gulabivala K. Effect of exposing dentine to NaOCl and calcium hydroxide on its flexural strength and elastic modulus. *Int Endod J*. 2001;34(2):113-119.
81. Doyon GE, Dumsha T, von Fraunhofer JA. Fracture resistance of human root dentin exposed to intracanal calcium hydroxide. *J Endod*. 2005;31(12):895-897.
82. Parirokh M, Torabinejad M. Mineral trioxide aggregate: a comprehensive literature review--Part I: chemical, physical, and antibacterial properties. *J Endod*. 2010;36(1):16-27.
83. Torabinejad M, Parirokh M. Mineral trioxide aggregate: a comprehensive literature review--part II: leakage and biocompatibility investigations. *J Endod*. 2010;36(2):190-202.
84. Mohammadi Z, Shalavi S, Soltani MK. Mineral trioxide aggregate (MTA)-like materials: an update review. *Compend Contin Educ Dent*. 2014; 35(8):557-561.
85. Roberts HW, Toth JM, Berzins DW, Charlton DG. Mineral trioxide aggregate material use in endodontic treatment: a review of the literature. *Dent Mater*. 2008;24(2):149-164.
86. Andreasen JO, Munksgaard EC, Bakland LK. Comparison of fracture resistance in root canals of immature sheep teeth after filling with calcium hydroxide or MTA. *Dent Traumatol*. 2006; 22(3):154-156.
87. Bortoluzzi EA, Souza EM, Reis MJ, Esberard RM, Tanomaru-Filho M. Fracture strength of bovine incisors after intra-radicular treatment with MTA in an experimental immature tooth model. *Int Endod J*. 2007;40(9):684-691.
88. Hatibović-Kofman S, Raimundo L, Zheng L, Chong L, Friedman M, Andreasen JO. Fracture resistance and histological findings of immature teeth treated with mineral trioxide aggregate. *Dent Traumatol*. 2008;24(3):272-276.
89. Tuna EB, Dinçol ME, Gençay K, Aktören O. Fracture resistance of immature teeth filled with BioAggregate, mineral trioxide aggregate and calcium hydroxide. *Dent Traumatol*. 2011;27(3):174-178.
90. Aksel H, Askerbeyli-Örs S, Deniz-Sungur D. Vertical root fracture resistance of simulated immature permanent teeth filled with MTA using different vehicles. *J Clin Exp Dent*. 2017; 9(2):e178-e181.
91. Guven Y, Tuna EB, Dinçol ME, Ozel E, Yilmaz B, Aktören O. Long-term fracture resistance of simulated immature teeth filled with various calcium silicate-based materials. *Biomed Res Int*. 2016;2863817.
92. Schmoldt SJ, Kirkpatrick TC, Rutledge RE, Yaccino JM. Reinforcement of simulated immature roots restored with composite resin, mineral trioxide aggregate, gutta-percha, or a fiber post after thermocycling. *J Endod*. 2011;37(10):1390-1393.
93. Sawyer AN, Nikonov SY, Pancio AK, et al. Effects of calcium silicate-based materials on the flexural properties of dentin. *J Endod*. 2012;38(5):680-683.
94. Elnaghy AM, Elsaka SE. Fracture resistance of simulated immature teeth filled with Biodentine and white mineral trioxide aggregate - an *in vitro* study. *Dent Traumatol*. 2016;32(2):116-120.
95. Moazami F, Sahebi S, Jamshidi D, Alavi A. The long-term effect of calcium hydroxide, calcium-enriched mixture cement and mineral trioxide aggregate on dentin strength. *Iran Endod J*. 2014;9(3):185-189.
96. Forghani M, Bidar M, Shahrami F, et al. Effect of MTA and Portland cement on fracture resistance of dentin. *J Dent Res Dent Clin Dent Prospects*. 2013;7(2):81-85.

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